



Messenger

Cyclosporiasis: Increase in Domestic Cases

The California Department of Public Health (CDPH), in coordination with the Centers for Disease Control and Prevention (CDC), has informed California local health jurisdictions of a recent national increase in reported laboratory-confirmed cyclosporiasis cases this summer. CDPH is closely monitoring cyclosporiasis cases in California and has requested enhanced surveillance for domestically-acquired cyclosporiasis.

As of August 1, the CDC received reports of 899 cyclosporiasis cases among persons without a history of international travel and with illness onset on or after May 1. Reports are from 31 states, including California. These cases include 20 clusters, none in California.

In California, CDPH has received reports of 39 cyclosporiasis cases within this same time period. Of these, 15 case-patients report no history of international travel and eight are pending travel history information. The 15 case-patients with domestically-acquired cyclosporiasis were reported by eight local health departments, mostly in southern California. At this time, no common exposures or specific vehicles of interest have been identified.

Currently, fresh basil from Mexico has been linked to 132 cyclosporiasis cases from 11 states: Connecticut, Florida, Georgia, Iowa, Massachusetts, Minnesota, New York, Ohio, Rhode Island, South Carolina, and Wisconsin. As a result, Siga Logistics de RL de CV of Mexico has [voluntarily recalled implicated lots of basil](#). No California cases have been linked to this product to date.

New Herpes Virus Test: HSV 1, 2 Amplification

On September 1, the County of San Luis Obispo Public Health Laboratory will initiate molecular amplification testing for herpes virus type 1 and 2, and will cease testing by cell culture (herpes virus isolation, test 6010) and direct fluorescent antibody typing (herpes DFA typing, test 6015).

This new test offers improved sensitivity (from 80-85% to 95%) with no loss in specificity (98-100%). The test turnaround time is approximately two days with no delay in reporting due to culture technique or typing.

With this change, the correct specimen will be a swab in Aptima multitest swab transport medium (STM), **not** viral transport medium (VTM).

[Download full specimen collection and transport instructions](#)



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SPECIMEN COLLECTION & TRANSPORT INSTRUCTIONS

Herpes Swab Collection Kit - For Amplification testing

SPECIMEN STORAGE & TRANSPORT

Temperature: Refrigerated or Room Temperature (2°C - 30°C / 35.6°F - 86°F)
Maximum Holding Time: 36 days
Exceptions: If needed store specimen at -20°C to -70°C for up to 36 days

KIT STORAGE

Temperature: room temperature (15°C - 30 °C / 59 °F - 86 °F)
Expiration: kit must be used prior to expiration date listed on kit label

ACCEPTABLE SPECIMEN TYPES

Swab of anogenital or oral lesion: swab in Aptima® Multitest Swab Specimen Collection Kit (STM).

TEST ORDER OPTIONS FOR THIS KIT

650: Herpes Virus 1 and 2 Amplification

REJECTION CRITERIA

- Holding time greater than 36 days
- Specimen container not labeled
- Specimen container has leaked or is not sealed properly
- Swab specimens that are missing the swab, contain multiple swabs, or contain the wrong swab

SPECIMEN COLLECTION

Optimal specimen collection time is within the first 3 days after appearance of the vesicular lesion. Prior to specimen collection, patients should avoid topical treatments as these may reduce virus yield.

1. Swab affected area:
 - a. **Vesicular Lesion:** Wipe vesicle with saline. Disrupt vesicle and collect fluid with swab. With same swab, collect cells from the base of the lesion.
 - b. **Non-Vesicular Lesion:** Pre-moisten swab with saline. Collect cells from base of the lesion.
2. Place swab into Aptima Multitest Swab Transport Medium (STM).
3. Place the collection tube inside the zippered portion of the biohazard specimen bag and seal tightly.
4. Fill out the requisition form completely. Fold it in half once (no staples please) and place in the outer pocket of the biohazard specimen bag (not inside the zippered pouch with the specimen).
5. Transport to the laboratory after collection. Specimen is stable in STM for 36 days.

Questions?

Please contact the Laboratory Director at 805-781-5512 or
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